

November 30, 2017

Certificate no.: SE-EE-171201-TE

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Abstract

A new superior Green Fluorescent Protein (GFP) mutant ~~known as~~ ~~called~~ superfolder GFP (sfGFP) ~~is was reported to be~~ more soluble, ~~folds faster~~ ~~folding~~, and ~~the brightest of~~ ~~brighter than~~ ~~the known~~ ~~any known~~ GFP mutants. This study aims to create a codon adapted sfGFP tag (TtsfGFP) ~~to that could~~ be used simultaneously ~~for~~ ~~in~~ protein localization ~~and~~ ~~as well as~~ affinity purification studies ~~in~~ *T. thermophile*. *In vivo* fluorescence spectroscopic analyses of a codon adapted and 6XHis tagged TtsfGFP cassette carrying clones ~~exhibited~~ ~~showed that they have~~ ~2-4 fold ~~increase in~~ fluorescence emission ~~as~~ compared ~~to~~ ~~with~~ control groups at 3rd hours. Fluorescence ~~et~~ microscopy results also revealed that TtsfGFP reaches ~~a~~ ~~maximum~~ emission ~~maxima~~ at 100 min, which is much earlier than EGFP and sfGFP controls ~~with~~ ~~at~~ ~240 min. ~~To test the affinity/localization dual tag features,~~ ~~a~~ *Tetrahymena* ATP dependent DNA ligase domain containing hypothetical gene (H) was cloned into the 3rd end of 6XHis-TtsfGFP, ~~in order to assess the affinity/localization dual tag feature~~. Hence, ~~ff~~ fluorescence ~~et~~ microscopic analysis of the 6XHis-TtsfGFP-H clone ~~confirmed~~ ~~revealed~~ its localization ~~into~~ the macro~~nucleus~~- and micronucleus of vegetative *T. thermophila*. Affinity ~~purifiability~~ ~~purification~~ of TtsfGFP and TtsfGFP-H with Ni-NTA beads was ~~furthermore~~ confirmed using SDS-PAGE and Western blot ~~analyses~~ ~~experiments~~. These results indicate that ~~the~~ 6XHis-TtsfGFP tag can be used as a dual tag for protein localization and affinity purification studies ~~in~~ *T. thermophila*.